Development of a novel RNA-based microsatellite stable predictive response signature (MSS-PRS) to identify MSS colorectal cancer (CRC) patients with a microsatellite instability-high (MSI-H) molecular phenotype GenèCentric 1040 Pappan KL, Mayhew GM, Shepherd JH, Guo J, Beebe KD, Eisner JR, Milburn MV

BACKGROUND

- MSI-H and mismatch repair deficient (dMMR) tumors are associated with favorable immune checkpoint inhibitor (ICI) responses (André et al., 2020), but up to 95% of CRC tumors are microsatellite stable (MSI-low; designated MSS hereafter)/proficient MMR (pMMR) leading to poorer prognosis and treatment outcomes than MSI-H/dMMR patients (Le et al., 2015, Ribic, et al., 2003).
- In MSS/pMMR metastatic CRC, multiple combination therapies are being investigated without biomarkers to guide therapy (Lizardo et al., 2020, Pecci et al., 2021).
- Therefore, we developed an RNA-based MSS predictive response signature (MSS-PRS) that selects tumors not identified with conventional MSI testing but have molecular characteristics consistent with microsatellite instability, making them a potential target for ICI.

METHODS

- MSS-PRS was developed using the TCGA CRC cohort (COAD; n=268). Training labels were assigned using the MSI Mantis score and mutation status from the top 8 genes with reported MSI association (i.e., 'training label genes'; Li et al., 2020, Sorokin et al., 2021).
- "Activated" samples had both one or more of the training label gene mutations and an MSI Mantis score \geq 0.4 (i.e., MSI-H), and "non-activated" samples were wild type for all training label genes and had an MSI Mantis score < 0.4. All other samples were designated "ambiguous" and excluded from training.
- Two thirds of the non-ambiguous samples were assigned to the classifier training set and all remaining samples were assigned to the test set. Using ClaNC software (Dabney, 2006) and cross-validation in the training set, a nearest centroid classifier was developed from a set of high mean, high variance candidate genes to select an optimal gene set to separate activated (PRS score > 0) and non-activated (PRS score < 0) groups. Classifier performance was evaluated in the test set as well as 4 additional separate CRC RNA-seq datasets accessed through GEO (GSE24551 and GSE39084) or cBioPortal (coad_silu_2022 and coad_cptac_2019).

RESULTS

The MSS-PRS contained 112 genes. Cross validation for its ability to correctly call MSI-H samples showed high agreement and exhibited comparable performance in both the TCGA COAD test and train set. Further, MSS-PRS produced a similar distribution in four additional CRC cohorts (Figure 1), validating its consistency in classifying colon adenocarcinoma samples.

Figure 1: Similar Distribution of MSS-PRS Scores between MSI-H and MSS **Tumors Across Five Independent Colon Cancer RNA-seq Datasets**



- The biological basis of the MSS-PRS was explored by dividing TCGA COAD into 4 groups defined by the intersection of MSI Mantis (MSI-H or MSS) and MSS-PRS (+ or -): Groups A, B, C, and D (Figure 2).
- Heat map display of the top 256 differentially expressed genes between Groups A, B, and C showed a clear distinction between Groups A and C, with Group B having an intermediate phenotype like Group A (Figure 2, left panel).







MMR genes (MLH1, MLH3, MSH2, MSH3, MSH6, PMS1, PMS2) were examined in cBioPortal to address whether MSS tumors harbored MMR alterations (Figure 4, left panel). Of the MMR genes, only MLH1 was included as a training label for MSS-PRS.

GeneCentric Therapeutics, Inc., Durham, NC

Figure 2: Differentially Expressed Genes Distinguish MSI-H/MSS-PRS (+) and MSS/MSS-PRS (-); MSS-PRS Signature Enriched with Immune Genes

Protein-protein interaction network mapping in String (Szklarczyk et al., 2023; https://string-db.org/) revealed a core set of immune genes related to chemotaxis, natural killer cells, and T-lymphocytes as well as several unlinked oneoff genes (Figure 2, right panel).

Figure 3: Cluster Analysis of Common Oncogenic Mutations in TCGA COAD by MSS-PRS and CMS

- Some gene alterations (e.g., BRAF) were more prevalent in MSI-H/MSS-PRS (+) tumors (Group A) whereas others (e.g., TP53, APC) were more frequent in MSS (Groups B and C) regardless of MSS-PRS status (**Figure 3**).
- TMB values were predominantly Hiah associated with MSI-H tumors.
- The Consensus Molecular Subtype 1 (CMS1) accounted for about two-thirds of MSI-H tumors whereas CMS2 was most prevalent in MSS Group C and about half the tumors of MSS Group B were CMS4 and the remainder was mixed.
- The mutation status and TMB of MSS Group B and C tumors were similar at the aggregate level, but notable differences were observed for CMS classification and KRAS and BRAF alterations. Group D, MSI-H/MSS-PRS (-), could not be characterized because it only contained 2 tumors.

Figure 4: MMR Gene Alterations More Common in MSI-H/MSS-PRS (+) than MSS Tumors and Immune Expression Profiles Separate MSS-PRS (+) and **MSS-PRS (-)**

MSI and MSS-PRS	Total,	MMR	MMR
Status	n	alt+ (%)	alt– (%)
MSI-H, MSS-PRS(+)	56	27 (48)	29 (52)
MSS, MSS-PRS(+)	98	12 (12)	86 (88)
MSS, MSS-PRS(-)	112	5 (4)	107 (96)
MSI-H, MSS-PRS(-)	2	0 (0)	2 (100)



Figure 5: T-cell Receptor (TCR) Repertoires Independently Confirm Immune Differences Between MSS-PRS (+) and MSS-PRS (-)



SUMMARY AND CONCLUSIONS

- them a potential target for ICI.

References

André T et al. N Engl J Med. 2020 Dec 3;383(23):2207-2218. Dabney AR. Bioinformatics. 2006 Jan 1;22(1):122-3. Le DT et al. N Engl J Med. 2015 Jun 25;372(26):2509-20. Li L et al. Comput Struct Biotechnol J. 2020 Mar 19; 18:668-675. Lizardo DY et al. Biochim Biophys Acta Rev Cancer. 2020 Dec;1874(2):188447. Pecci F et al. Curr Treat Options Oncol. 2021 Jun 10;22(8):69. Ribic CM et sl. N Engl J Med. 2003 Jul 17;349(3):247-57. Sorokin M et al. Front Mol Biosci. 2021 Nov 23; 8:737821. Szklarczyk D et al. Nucleic Acids Res. 2023 Jan 6;51(D1):D638-D646.

As expected, Group A (MSI-H/MSS-PRS+) carried the most MMR alterations with 48% of tumors having at least one alteration. While Group B (MSS/MSS-PRS+) tumors had more MMR alterations, 12%, than Group C (MSS/MSS-PRS-), 4%, these differences did not explain the difference in molecular phenotype between these two MSS groups for the majority of tumors (**Figure 4**, **left panel**)

In contrast, gene expression-based immune profiles in MSS tumors called activated (Group B) were markedly more like MSI-H tumors (Group A) compared to MSS tumors called non-activated (Group C; Figure 4, right panel).

Examination of T cell receptor (TCR) α/β repertoires confirmed differences in adaptive immunity between MSS-PRS+ (Groups A and B) and MSS-PRS- (Group C; Figure 5). TCR clonal expansion is an epigenomic event, thus the differences in TCR repertoire offer an independent indication that the immune molecular state of Groups A and B are more similar to each other than to Group C.

Herein we described the development of the novel MSS-PRS that selects tumors not identified with conventional MSI testing (e.g., MSI-H/dMMR) but have molecular characteristics consistent with microsatellite instability, thus making

Based upon these initial findings, further development of the MSS-PRS and its clinical validation as a tool to select patients with MSS tumors who may benefit from ICI-containing treatment regimens is warranted.

Efforts are ongoing to develop the MSS-PRS as a novel RNA-based diagnostic test and further evaluate it in both retrospective and prospective clinical studies. Additionally, to expand the potential utility of MSS-PRS further, a liquid biopsy analog of this signature is under development.